



Immune response of *Salmonella enteritidis* antigens in rabbits

Afaf, A. Yousif and ¹Sabaa H. H. Al-Mansory

Department of Internal and Preventive Medicine, College of Veterinary Medicine, Baghdad University, Iraq

¹Department of Internal and Preventive Medicine, College of Veterinary Medicine, Al-Qasem, Iraq

Abstract

This study aimed to examine the effect of two inactivated *Salmonella enteritidis* antigens (O and H Ag) on humoral and cellular immunity in rabbits. Twelve rabbits of matching age and weight were divided into 3 equal groups. The first group was injected subcutaneously with 1 ml of O Ag, the second group was injected similarly with H Ag and the third group, (control) was injected with 1 ml phosphate buffer saline (PBS). Blood samples were collected at 2, 4, and 8 weeks after injections. Two-way analysis of variance was used to test the significance of the effects of treatments and period on hemagglutination test and delayed type hypersensitivity skin test (DTH). Eight groups of rabbits (4 rabbits per group) were used to determine the lethal dose 50 (LD₅₀). All rabbits of the treated and control groups were challenged with 4 LD₅₀ (8×10¹⁰) of virulent *Salmonella enteritidis*. Passive hemagglutination test showed significant increase in antibody titers in the two experimental groups compared with the control. Cellular immunity estimated by DTH gave higher response, while control group did not show any reaction. The post challenge with LD₅₀ of *S. enteritidis* revealed that all rabbits in control group suffered from severe clinical signs of salmonellosis and died within 3-4 days. The present study indicated that injection of O and H Ag improved the immune response in rabbits.

Keywords: *Salmonella enteritidis*, O and H Antigens, Humoral immunity, Cellular Immunity

Introduction

Salmonella enteritidis remains one of the main causes of food-borne disease. It is considered as the most important pandemic zoonotic disease that spreads under natural conditions (Rodrigue et al., 1990). *Salmonella enterica* serovar *enteritidis* has emerged during the last 20 years as the major causative agent of food-borne gastro-enteritidis in humans (Toyofuku, 2008). There is a need to develop new vaccines and therapeutics (Mastroeni et al., 2001; Mastroeni and Grant, 2011) due to the difficulty in the prevention of salmonellosis by implementation of hygiene measures. In rabbits, salmonellosis is characterized by septicemia and rapid death with diarrhea and abortion. Mortality is the highest in young rabbits and pregnant does. Bacteria are shed in the faeces of carrier rabbits and clinically ill animals (Patton et al., 2008). The rabbit can be used as a model for diarrheal disease and sequel associated with salmonellosis (Hanes et al., 2001).

Salmonella are Gram-negative, flagellated, facultative anaerobic bacteria. The bacilli possess three major antigens: H (flagella antigen), O (somatic antigen) and Vi (virulence antigen) possessed by only a serovars (Duguid et al., 1989; Rubin and Weinstein, 2004). Ibebuike et al. (2008) showed that O and H antigens have the possibility of producing high *Salmonella* antisera against antigens. Also, Nalbantsoy et al. (2010) isolated and purified O and H antigens from *S. enteritidis*.

This study was designed to evaluate the humeral and cellular immune response in rabbits following exposure to O and H antigens of *S. enteritidis*.

Materials and Methods

Salmonella enteritidis was isolated from Iraqi goats by culturing on different selective media and by biochemical tests (Quinn et al 2004; AL-Shemmari, 2008). The bacterial confirmation was done in *Salmonella* Centre in Baghdad, Ministry of Health.

Corresponding author: A. Yousif, Department of Internal and Preventive Medicine/College of Veterinary Medicine, Baghdad University

tests was prepared according to Mitov et al. (1992). Briefly, a bacterial suspension of *S. enteritidis* obtained from overnight agar culture was sonicated at 50 minute intervals in a water cooled sonicator oscillator at 40 MHZ per second and the homogenate was centrifuged twice by using a cooling centrifuge at 8000 rpm per 30 minutes each time to remove cellular debris. The supernatants were passed through a 0.22 µm Millipore filter and stored at (-20°C) until used. Protein content was determined by biuret protein assay.

The somatic antigen (O Ag) was prepared by heating bacterial suspension of *S. enteritidis* on water bath at 100°C for 30 minutes. The flagller antigen (H Ag) was prepared by adding 3% formaldehyde solution to the bacterial suspension of *S. enteritidis* and left overnight (Smith et al., 1984). The two antigens were tested prior for sterility and safety before use according to OIE (2004).

To evaluate the efficacy of the prepared antigens, 12 adult healthy rabbits aged 6 to 8 months were selected. All rabbits had negative faecal bacteriological culture for salmonella. They were reared in separate cages in the Animal House of Veterinary College, University of Baghdad. The animals were divided equally into three groups. The first group (Immunized with O Ag) was injected twice at two weeks intervals S/C with the prepared O antigen at a dose of 1 ml containing 1×10^8 CFU/ml. The second group was injected with the prepared H antigen at same dose. The third group (control) was injected S/C with 1 ml of PBS. Blood samples were collected from all groups at 2nd, 4th and 8th week post-injection. Sera were separated and stored at -20°C.

The humoral immunity was evaluated by passive haemagglutination test (PHA) as described by Boyden (1951). DTH-skin test as described by Hudson and Hay (1980). Briefly, 0.1 ml of soluble antigen of *S. enteritidis* was injected intradermally in the right flank of the immunized and control groups while the left flank was injected by 1 ml of sterile PBS (pH=7.2). The

thickness of skin was measured by vernier calliper before injection and at 24, 48 and 72 hours post injection.

The lethal dose 50 (LD₅₀) was estimated by using eight fold dilution for counting bacteria by viable bacterial plate count method (Quinn et al., 2004). Thirty two healthy rabbits of both sexes were divided into (8) groups (4 per group). Seven groups of rabbits were injected subcutaneously with 1 ml of calculated CFU diluents, and the eighth group was considered as a control group injected with PBS. All groups were monitored for 30 days to calculate the total live and dead rabbits, and also to estimate the LD₅₀ according to Reed and Muench (1938). After 8th weeks, all immunized and control rabbits were challenged intra-peritoneally with 4 LD₅₀ virulent *S. enteritidis* to evaluate the efficacy of prepared the antigens in inducing immune response.

Statistical Analyses

Using statistical package for social science (SPSS) version 13.0, two-way analysis of variance was conducted to test the significance of effects of groups and periods post injection on the examined traits. The statistical differences among means of the different treatments were tested By Duncan's multiple range test.

Results

The PHA test showed a significant increase ($P < 0.05$) in antibody titre in the groups injected with O and H Ag compared to the control group. Also, the titre was significantly high on 4th weeks compared to 2nd and 8th week (Table 1). DTH tests indicated that the values were significantly high in the experimental groups compared to the control group. Again, the thickness (mm) increased ($P \leq 0.05$) after 48 hours and then returned to normal (Table 2&3). The results of mortality are shown in Table 4.

Table 1: Antibodies titre (Mean ± SE) of the experimental and control groups by PHA test

Weeks of injection	O Ag injected group	H Ag injected group	Control group
2 nd	^b 38.00±6.25 ^A	^c 28.00±4.00 ^A	0.00 ^B
4 th	^a 88.00±24.00 ^A	^a 72.00±20.13 ^A	0.00 ^B
8 th	^b 48.00±9.23 ^A	^b 64.00±8.23 ^A	0.00 ^B

^{a-c} means in the same column with different (small letter) superscripts differed significantly at $P < 0.05$; ^{A-B} Means in the same row with different (capital letter) superscripts differed significantly at $P < 0.05$

Table 2: Mean skin thickness (Mean ± SE) of experimental and control groups in DTH test

Periods after injection of soluble antigen	Diameter of skin thickness (mm)		
	O Ag injected group	H Ag injected group	Control group
24 hours	^b 0.42±0.06 ^A	^b 0.52 ± 0.08 ^A	^a 0.27±0.07 ^B
48 hours	^a 0.70 ± 0.08 ^A	^a 0.90 ± 0.16 ^A	^a 0.40± 0.08 ^B
72 hours	^b 0.50 ±0.168 ^A	^b 0.52 ± 0.08 ^A	^a 0.20±0.04 ^B

^{a-c} Means in the same column with different (small letter) superscripts differed significantly at $P < 0.05$; ^{A-B} Means in the same row with different (capital letter) superscripts differed significantly at $P < 0.05$

Table 3: Mean skin redness (Mean ± SE) of experimental and control groups in DTH test

Periods after injection of soluble antigen	Diameter of skin redness(mm)		
	O Ag injected group	H Ag injected group	Control group
24 hours	^b 0.67 ± 0.13 ^A	^b 0.42 ± 0.10 ^B	0.00 ^C
48 hours	^a 1.00 ± 0.05 ^A	^a 0.97 ± 0.04 ^A	0.00 ^C
72 hours	^c 0.37 ± 0.02 ^A	^b 0.42 ± 0.06 ^A	0.00 ^C

^{a-c} means in the same column with different (small letter) superscripts differed significantly at P<0.05; ^{A-B} means in the same row with different (capital letter) superscripts differed significantly at P<0.05

Table 4: Mortality rate (%) of LD₅₀ of *S. enteritidis* in rabbits

Rabbits group	*Calculated dose	Alive	Dead	Total	Alive	Total	Dead	Percent mortality
1	2×10 ¹²	0	4	0	10	100 %		
2	2×10 ¹¹	1	3	1	6	81 %		
3	2×10 ¹⁰	2	2	3	3	50 %		
4	2×10 ⁹	3	1	6	1	18 %		
5	2×10 ⁸	4	0	10	0	0 %		
6	2×10 ⁷	4	0	14	0	0 %		
7	2×10 ⁶	4	0	18	0	0 %		
8	PBS	4	-	-	-	0%		

No. of rabbits in each group = 4, Total No. of rabbits = 32, *The dose calculated as (cells).

Clinical signs post challenge

All rabbits injected with O and H Ag and control group challenged with 4 LD₅₀ (4×2×10¹⁰) 8 weeks post immunization exhibited moderate elevation in the body temperature, which persisted for 2-3 days with mild signs of illness and depression without diarrhoea and returned to normalcy within 7 days.

The control group also exhibited the clinical signs post challenge. These signs included listlessness, anorexia, severe diarrhoea, fever, rough coat, hunched posture and crowding near the water supply. Increase in the respiration rate, abortion of the pregnant does, severe dehydration and recumbency appeared at the later stage and death occurred within 3 to 5 days after the challenge.

Discussion

Since *Salmonella* organisms are ubiquitous pathogens of human and animal species, an understanding of events during the immune response is of paramount importance in developing an effective prophylactic agent (Collins, 1974). The important role of antibody producing B cell in protection against salmonellosis has been reported in many studies (Smith et al., 1993; Lindberg et al., 1993; Mastroeni et al., 2000). In the current study, inoculation of rabbits with O and H Ag of *S. enteritidis* resulted in stimulation of significant high titres of antibody in the experimental groups compared with control. This is in agreement with George et al. (1985), who reported that immunization of rabbits with H and O. *Salmonella* antigens resulted in high yield of antisera titre. Weidans et al. (1964) recorded that the rabbits injected with bacterial cells or isolated somatic antigen of *S. enteritidis* and *Escherichia coli* produced higher cellular and humoral immune response. Mittrucker and

Kaufmann (2000) mentioned that the response to *S. typhimurium* involves both T and B cell-mediated immunity which are important for control of primary infection and protection against secondary infection attack.

In the present study, the first group inoculated with O Ag evoked antibody response, and this is compatible with Ibebuikwe et al. (2008) and Saxon et al. (1986) who demonstrated the possibility of producing high *Salmonella* antisera by inoculating rabbits with salmonella antigens. Also, the second group immunized with H Ag revealed significantly higher titres of antibody compared with the control group. The significance of such immune response has been highlighted in several studies (Stocker and Newton, 1994; Ibebuikwe et al., 2008).

Many investigations have concluded that cellular immunity is the primary mechanism of protection against salmonellosis, especially when vaccines are employed (Mastroeni et al., 1993). Our study showed significant increase in the antibody titre against O and H antigen, indicating the possibility of the stimulation of cellular immunity. This result is in agreement with reports which recorded cellular immune against attenuated or heat killed *S. typhimurium* or with outer membrane proteins or flagellin (Ogunniyi et al., 1994; Thatte et al., 1995; Cookson and Bevan 1997; Aderem and Ulevitch, 2000).

In this investigation, delayed type hypersensitivity skin test showed significantly higher value in experimental animals. This result is in agreement with the study of Strindelius et al. (2002) who used delayed-type hypersensitivity – skin test as a measure of cellular immunity in mice immunized with different types of *S. enteritidis* antigens and record a significant increase in ear thickness of all immunized groups. Also, Mitov et al. (1992) detected positive DTH test in immunized

mice with salmonella and their results revealed development of long lasting cell mediated immunity that showed striking correlation to the protection.

The immunized groups in our study resisted the effect of lethal challenge and all were live after immunization with O and H antigens and due to its ability to reduce the appearances of severe clinical signs of salmonellosis while the control group showed severe clinical signs of salmonellosis and died within 3-4 days after challenge. These results are in agreement with Uchiya et al. (1991) and Karasova (2009) who reported that of mice with *S. enteritidis* induced strong cellular immunity and resisted the lethal challenge. Also, Simon et al. (2011) reported that the mice immunized with flagellin alone, or flagellin conjugates were protected from lethal challenge with wild-type *S. enteritidis*.

In conclusion, immunization of rabbits with O and H *S. enteritidis* antigens induced humoral immune response as detected by the studied parameters.

References

- Aderem, A. and Ulevitch, R.J. 2000. Toll-like receptors in the induction of the innate immune response. *Nature*, 406:782-787.
- AL-Shemmari, I.G.M. 2008. Epidemiological study of *Salmonella spp.* isolated from goat in some provinces in middle of Iraq. Master thesis, Vet. Med. College/University of Baghdad.
- Boyden, S.V. 1951. Fixation of bacterial products by erythrocytes treated with tannic acid and subsequent hemagglutination by anti-protein sera. *Journal of Experimental Medicine*, 93: 107.
- Collins, F.M. 1974. Vaccines and cell mediated immunity. *Bacteriological Review*, 38: 371.
- Cookson, B.T. and Bevan, M.J. 1997. Identification of a natural T cell epitope presented by *Salmonella*-infected macrophages and recognized by T cells from orally immunized mice. *Journal of Immunology*, 158:4310-4319.
- Duguid, J.P., Collee, J.G., Fraser, A.G. and Marmion, B.P. 1989. Practical Microbiology 13th (Ed.). Churchill Livingstone London. P: 481.
- George, F.I., Graham, H.F., Mary, J.L. and Reta, A.W. 1985. Production of potent *Salmonella* H antisera by immunization with polymeric flagellins. *Journal of Clinical Microbiology*, 22: 347-351.
- Hanes, D.E., Robl, M.G., Schneider, C.M. and Burr, D.H. 2001. New Zealand white rabbit as a nonsurgical experimental model for *Salmonella enterica* gastroenteritis. *Infection and Immunity*, 69(10): 6523-6.
- Hudson, L. and Hay, F.C. 1980. Practical Immunology. 3rd (ed.). Black-Well Scientific Publications, Oxford London.
- Ibebuikel, C.C., Yah, S.C. and Eghafona, N.O. 2008. Production of high polyvalent antisera against *Salmonella*. *Scientific Research and Essay*, 3 (5): 204-208.
- Karasova, D. Sebkova, A., Vrbas, V., Avlickova, H., Sisak, F and Rychlik, I. 2009. Comparative analysis of salmonella enterica serovar enteritidis mutants with a vaccine potential. *Vaccine*, 27: 5265-5270.
- Lindberg, A.A., Segall, T., Weintraub, A. and Stocker, B.A.D. 1993. Antibody response and Protective against challenge in mice vaccinated intraperitoneally with a live aroA O4-O9 hybrid *S. dublin* strain. *Infection and Immunity*, 61: 1211.
- Mastroeni, P., Chabalgoity, J.A., Dunstan, S.J., Maskell, D.J. and Dougan, G. 2001. *Salmonella*: immune responses and vaccines. *Veterinary Journal*, 161:132-164.
- Mastroeni, P. and Grant, A.J. 2011. Spread of *Salmonella enterica* in the body during systemic infection: unraveling host and pathogen determinants. *Expert Review in Molecular Medicine*, 13: 12.
- Mastroeni, P., Simmons, C., Fowler, R., Hormaeche, C.E.; and Dougan, G. 2000. Igh-6(-/-) (B-cell-deficient) mice fail to mount solid acquired resistance to oral challenge with virulent *Salmonella enterica* serovar Typhimurium and show impaired Th1 T-cell responses to *Salmonella* antigens. *Infection and Immunity*, 68:46-53.
- Mastroeni, P., Villarreal-Ramos, B. and Hormaeche, C.E. 1993. Effect of late administration of anti-TNF α antibodies on a *Salmonella* infection in mouse model. *Microbial Pathogenesis*, 14: 473.
- Mitov, I., Denchev, V. and Linde, K. 1992. Humoral and cell-mediated immunity in mice after immunization with live oral vaccines of *S. typhimurium*: auxotrophic mutants with two attenuating markers. *Vaccine*, 10(1): 61-66.
- Mittrucker, H.W. and Kaufmann, S.H. 2000. Immune response to infection with *Salmonella typhimurium* in mice. *Journal of Leukocyte Biology*, 67:457-463.
- Nalbantsoy, A., Karaboz, I., Ivanova, R. and Deliloglu-Gurhan, I. 2010. Isolation and purification of O and H antigens from *Salmonella* Enteritidis as diagnostic tool. *Annals of Microbiology*, 60(3): 565-571.
- Ogunniyi, A.D., Manning, P.A. and Kotlarski, I. 1994. A *Salmonella enteritidis* 11RX pilin induces strong T-lymphocyte responses. *Infection and Immunity*, 62: 5376-5383.
- OIE 2004. Manual of diagnostic tests & vaccines for Terrestrial Animal. Fifth Edition. 1018.
- Patton, N.M., Hagen, K.W., Gorham, J.R. and Flatt, R.E. 2008. Domestic Rabbits: Diseases and Parasites. A Pacific Northwest Extension Publication, *Oregon Idaho Washington*, PNW 310-E.

- Quinn, P.J., Carter, M.E., Markey, B. and Carter, G.R. 2004. *Clinical Veterinary Microbiology*. 6th (ed.). Mosby-Wolf, London.
- Reed, L.J. and Muench, H. 1938. A simple method of estimating fifty percent end point. *American Journal of Hygiene*, 27(16): 8739-8744.
- Rodrigue, D.C., Tauxe, R.V. and Rowe, B. 1990. International increase in *Salmonella enteritidis*: a new pandemic? *Epidemiology and Infection*, 105:21–27.
- Rubin, R.H. and Weinstein, L. 2004. *Salmonellosis: Microbiologic, pathologic and clinical features*. Stratton Intercontinental Medical Book Corp. New York. P: 385.
- Saxén, H., Nurminen, M., Kuusi, N., Svenson, S.B. and Mäkelä, P.H. 1986. Evidence for the importance of O antigen specific antibodies in mouse-protective *Salmonella* outer membrane protein (porin) antisera. *Microbial Pathogenesis*, 1:433-441.
- Simon, R., Tennant, S.M., Wang, J.Y., Schmidlein, P.J., Lees, A., Ernst, R.K., Pasetti, M.F., Galen, J.E. and Levine, M.M. 2011. *Salmonella enterica* Serovar *Enteritidis* Core O Polysaccharide Conjugated to H:g,m Flagellin as a Candidate Vaccine for Protection against Invasive Infection with *S. enteritidis*. *Infection and Immunity*, 79(10): 4240-4249.
- Smith, B.P., Dilling, G.W., Roden, L.D. and Stocker, B.A. 1993. Vaccination of calves with orally administered aromatic-dependent *Salmonella dublin*. *American Journal of Veterinary Research*, 54: 1249-1255.
- Smith, B.P., Reina-Guerra, M., Hoiseth, S.K., Stockert, A.B., Habasha, F., Johnson, E. and Merritt F. 1984. Aromatic dependent *S. typhimurium* as modified the vaccine for calves. *American Journal of Veterinary Research*, 45: 181-89.
- Stocker, B.A. and Newton, S.M. 1994. Immune responses to epitopes inserted in *Salmonella* flagellin. *International Review of Immunology*, 11:167-178.
- Strindelius, L., Lena, D.W. and Ingvar, S. 2002. Extracellular Antigens from *Salmonella enteritidis* Induce Effective Immune Response in Mice after Oral Vaccination. *Infection and Immunity*, 70(3): 1434–1442.
- Thatte, J. Rath, S. and Bal, V. 1995. Analysis of immunization route-related variation in the immune response to heat-killed *Salmonella typhimurium* in mice. *Infection and Immunity*, 1995 Jan; 63(1): 99-103.
- Toyofuku, H. 2008. Epidemiological data on food poisonings in Japan focused on *Salmonella*, 1998-2004. *Food Additives and Contaminants*, 26:1-9.
- Uchiya, K., Nikai, T. and Sugihara, H. 1991. Difference in the protection against infection with different challenge strains of *Salmonella Enteritidis* by killed vaccine. *Kansenshogaku Zasshi*, 65:1411–1418.
- Weidanz, W.P., Jackson, A.L. and Landy, M. 1964. Some aspects of antibody response of rabbits to immunization with Enterobacterial somatic antigens *Proceedings of the Society for Experimental Biology and Medicine*, 116: 832.